

## Total Synthesis of Sulfobacin A (Flavocristamide B)

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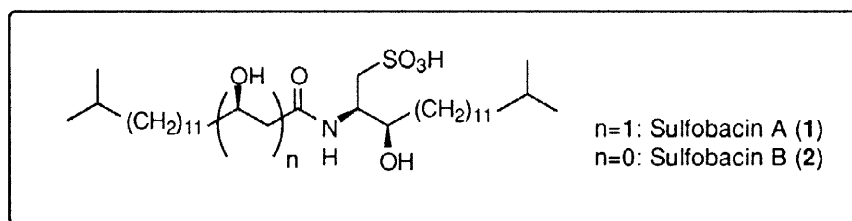
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**Abstract:** Sulfobacin A (1), a novel von Willebrand factor receptor antagonist isolated from the culture broth of *Chryseobacterium* sp. NR 2993, was efficiently synthesized for the first time.

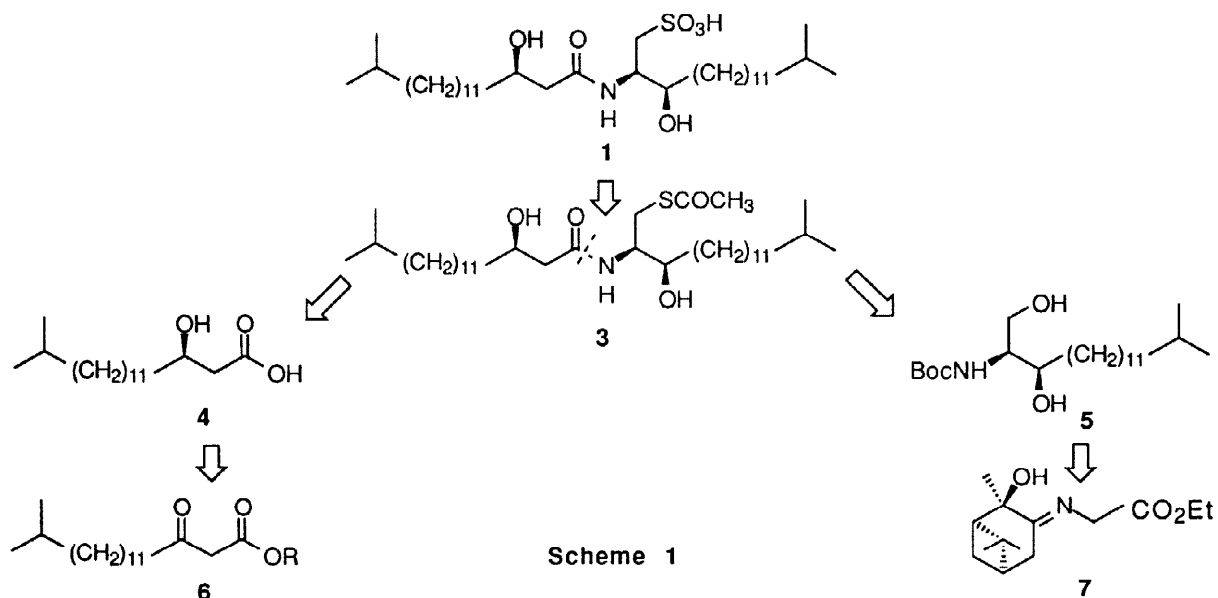
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Sulfobacins A (1) and B (2), novel von Willebrand factor (vWF) receptor antagonists, have been isolated by Kamiyama *et al.*<sup>1</sup> from the culture broth of *Chryseobacterium* sp. (*Flavobacterium* sp.) NR 2993 in a soil sample collected in Iriomote Island, Okinawa Prefecture, Japan. Sulfobacin A (1) was also isolated by Kobayashi *et al.*<sup>2</sup> as flavocristamide B from *Flavobacterium* sp. in the marine bivalve *Cristaria plicata* collected in Ishikari Bay, Hokkaido, Japan. Sulfobacins A (1) and B (2) inhibit the binding of vWF to the GPIIb/IX receptors in a competitive manner with IC<sub>50</sub>s of 0.47 and 2.2 μM, respectively.<sup>1a</sup> Furthermore, sulfobacin A (1) was found to have inhibitory activity against DNA polymerase α.<sup>2</sup> The structures are related to sulfonolipids having an aminosulfonic acid moiety and are analogous to sphingosine. The absolute configurations of the sulfobacins were determined using the modified Mosher method.<sup>1b</sup> We now wish to report the first total synthesis of sulfobacin A (flavocristamide B, 1) in an effective and stereoselective manner.

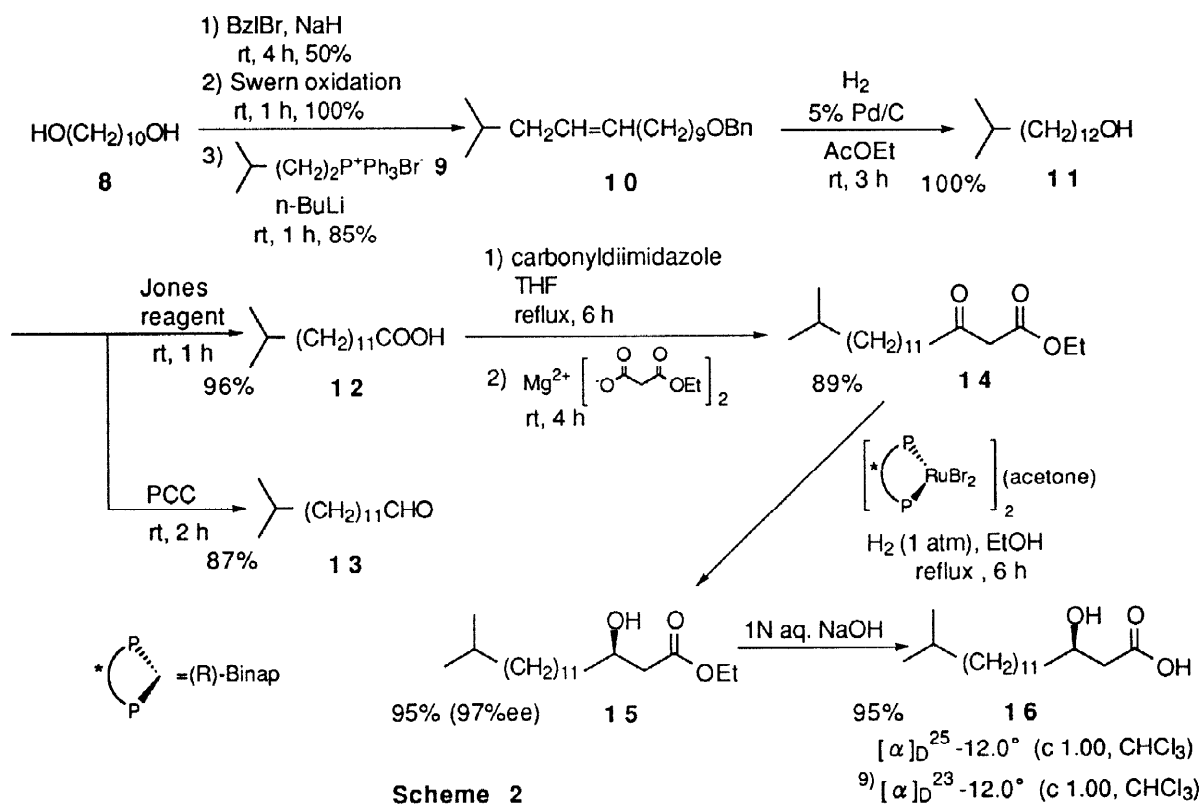


Sulfobacin A (1) could be prepared from the corresponding thioacetate 3, which could be constructed by coupling the left fragment 4 with the right fragment 5 using diethyl phosphorocyanidate (DEPC, (C<sub>2</sub>H<sub>5</sub>O)<sub>2</sub>P(O)CN)<sup>3</sup> as a coupling reagent. The left fragment 4 would be obtained through the asymmetric reduction of the corresponding β-keto ester 6. The right fragment would be constructed by the asymmetric aldol reaction using the Schiff base derived from (+)-2-hydroxy-3-pinanone ((+)-HyPN, 7).<sup>4</sup>

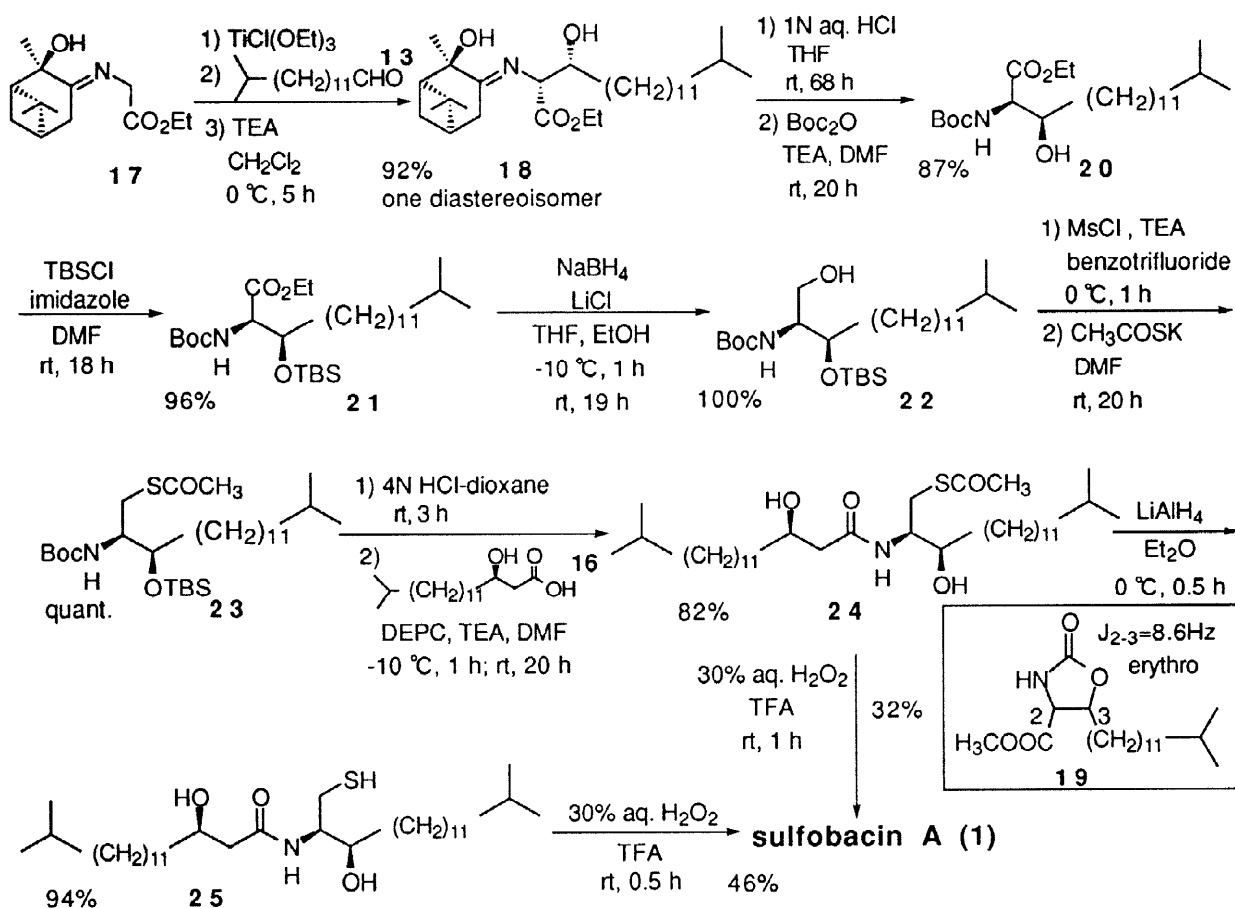
The synthesis of the left fragment started from 1,10-decanediol (8). After protection of one of the hydroxy groups of 8 with benzyl bromide,<sup>5</sup> the Swern oxidation followed by the Wittig reaction with the ylide from the phosphonium bromide 9<sup>6</sup> afforded the olefin 10. Reduction of the double bond and hydrogenolytic deprotection over a 5% Pd/C catalyst afforded the alcohol 11, which was converted to the carboxylic acid 12 with Jones reagent. The aldehyde 13 was also obtained from 11 by oxidation with PCC.



Reaction of the carboxylic acid **12** with carbonyldiimidazole followed by the magnesium enolate of the malonic acid half-ester yielded the  $\beta$ -keto ester **14**. The asymmetric hydrogenation of the  $\beta$ -keto ester **14** with chiral Ru(II) catalysts at atmospheric pressure according to Genêt's method<sup>7</sup> smoothly proceeded to give the  $\beta$ -hydroxy ester **15** in 95% yield with 97% ee,<sup>8</sup> which was converted to the  $\beta$ -hydroxycarboxylic acid **16** by alkaline treatment. The stereogenic center of **16** was revealed to be (*R*) according to the comparison of the specific rotation of that already reported.<sup>9</sup>



The right fragment was prepared from the aldehyde **13** and the chiral Schiff base of (+)-2-hydroxy-3-pinanone ((+)-HyPN, **17**) using the Solladié's methodology.<sup>10</sup> Thus, the asymmetric aldol reaction of the aldehyde **13** with the chiral titanium enolate generated from titanium chlorotriethoxide and the Schiff base **17** gave the erythro aldol adduct **18** in 92% yield as a single diastereoisomer. This adduct **18** was converted<sup>11</sup> to the oxazolidine **19** and the configuration was determined to be erythro by its <sup>1</sup>H NMR spectral analysis. The absolute configuration was determined using a modified Mosher method.<sup>12</sup> Removal of the chiral auxiliary with 1N aq. HCl, followed by treatment with Boc<sub>2</sub>O, afforded the hydroxy ester **20**. After protection of the hydroxy group with *tert*-butyldimethylsilyl (TBS) chloride, the ester **21** was reduced to give the primary alcohol **22**, which was converted to the thioacetate **23** via the mesylate. After deprotection of the thioacetate **23** with hydrogen chloride in dioxane, the coupling of the deprotected right fragment with the left fragment **16** was smoothly achieved with DEPC. The thioacetate **24** was subjected to pertrifluoroacetic acid oxidation to give sulfobacin A (**1**). Alternatively, the thioacetate **24** was reduced with LiAlH<sub>4</sub> to give the corresponding thiol which underwent the pertrifluoroacetic acid oxidation to yield sulfobacin A (**1**). The synthetic sulfobacin A ([α]<sub>D</sub><sup>18</sup> -31.6° (c 0.14, MeOH)) was identical with the natural one<sup>1</sup> ([α]<sub>D</sub><sup>24</sup> -35° (c 0.14, MeOH)) in every respect (IR, <sup>1</sup>H NMR, <sup>13</sup>C NMR spectra, and TLC). Thus we have completed the first total synthesis of sulfobacin A (**1**).



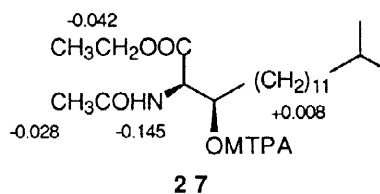
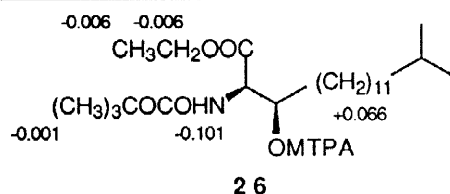
Scheme 3

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## References and Notes

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5. The dibenzyl derivative was also obtained in 7% yield.
6. The phosphonium bromide **9** was prepared from isoamyl bromide and triphenyl phosphine in acetonitrile at reflux temperature.
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8. Determined by  $^1\text{H}$  NMR spectral analysis of the corresponding MTPA ester derivative.
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11. The adduct **18** was converted to the oxazolidinone **19** by the removal of the chiral auxiliary with hydrogen chloride in dioxane, amino protection with 4-methoxybenzyl S-(4,6-dimethylpyrimidin-2-yl)-thiocarbonate (Z(OMe)-SDP)(75%), cyclization with 1*N* NaOH, and methylation with iodomethane (80% in 2 steps).
12. The absolute configuration was determined by the  $\Delta\delta(\delta_S-\delta_R)$  values (ppm) obtained from  $^1\text{H}$ -NMR spectral data for the MTPA esters **26** and **27** in  $\text{CDCl}_3$ .

$\Delta\delta(\delta_S-\delta_R)$  values(ppm) obtained from  $^1\text{H}$ -NMR spectral data in  $\text{CDCl}_3$



*Cf.* Ohtani, I.; Kusumi, T.; Kashima, Y.; Kakisawa, H. *J. Am. Chem. Soc.* **1991**, *113*, 4092.